Activation of peroxyl and molecular oxygen using *bis*-benzimidazole diamide copper (II) compounds

PAVAN MATHUR

Department of Chemistry, University of Delhi, Delhi 110 007 e-mail: pavanmat@yahoo.co.in

Abstract. New tetradentate *bis*-benzimidazole ligands have been synthesized and utilized to prepare copper (II) complexes. Some of these copper (II) complexes have been characterized structurally. The copper (II) in these complexes is found to possess varying geometries. A distorted octahedral geometry is found with a highly unsymmetrical bidentate nitrate group. An unusual polymeric one-dimensional structure is observed where copper (II) is in a distorted square pyramidal geometry with a monodentate nitrate ion, having long Cu-O bond, while a distorted triagonal bipyramidal geometry is found with two carbonyl O atoms and a Cl atom in the equatorial plane, and two benzimidazole imine N atoms occupy the axial position. These compounds are found to activate the cumylperoxyl group, and this has been utilized in the facile oxidation of aromatic alcohols to aldehydes, where they act as catalysts with large turnovers. The yields of the respective products vary from 32 to 65%. The role of molecular oxygen has been studied and an attempt has been made to identify the "active copper species". Activation of molecular oxygen has also been observed and has been used for oxidative dealkylation of a hindered phenol, producing di-butyl quinones with yields of 20–25% and 10–12 fold catalytic turnover. Dihydroxybenzenes and substituted catechols are also readily oxidized to the corresponding quinones, in oxygen-saturated solvents. Yields of 84% have been observed with 34-fold catalyst turnover, with di-t-butylcatechol. The activity of these copper (II) - bis-benzmidazolediamide compounds is reminiscent of the functioning of copper centres in galactose oxidase, tyrosinase and catechol oxidase.

Keywords. Copper(II); bis-benzimidazolediamide; galactose oxidase; catechol oxidase; tyrosinase.

1. Introduction

Oxidation reactions using molecular oxygen are of fundamental significance in living organisms. The ability of copper-containing metalloproteins to reversibly bind and activate dioxygen has been the focus of continued interest in bioinorganic chemistry. Tyrosinase is a copper-containing enzyme that contains two Cu(II) ions coupled via a magnetic exchange interaction. This coupled dinuclear Cu(II) site is also the resting state of the enzyme and is referred to as mettyrosinase.¹ The enzyme functions via a deoxy site which has two copper ions in the +1 oxidation state. An oxytyrosinase form is generated when the deoxy site reacts with molecular oxygen. The oxytyrosinase hydroxylates monophenols to diphenols and in turn produces the met-tyrosinase form containing two Cu(II) ions. It is the met-tyrosinase form that is then further involved in the oxidation of diphenols to quinones, regenerating the deoxy site. This activity of mettyrosinase is similar to that of catechol oxidase from sweet potatoes² while galactose oxidase (GO) is a fungal enzyme containing mononuclear copper ion, which

catalyses the oxidation of primary alcohols to aldehydes, coupled to the two-electron reduction of O_2 to $H_2O_2^{-3}$

In its catalytic cycle, a unique tyrosyl radical cofactor plays a vital role.⁴ The Cu(II) is located in a distorted square based pyramidal environment with a tyrosine in the apical position; Two histidines, Try-272, and an acetate ion form the N_2O_2 square planar geometry around the Cu(II) site.⁵

Further there is interest in the research devoted to ligands bearing an amido group as they act as fundamental building blocks in many biomolecules.⁶ A number of ligands based on salicylamide⁷ and pyridine mono and diamide⁸ have been reported which resist oxidative degradation and afford very stable complexes with a vide range of metal ions. Interest in the use of chelating ligands incorporating benzimidazole as models for active sites in several copper containing proteins has also grown rapidly since imidazole of histidines was identified as one of the coordinating ligands in many of the copper-containing proteins.

Moreover, the catalytic oxidation and oxygenation of organic substrates by biomimetic metal complexes is equally important. Such complexes provide an alternate route to cleaner organic synthesis using molecular oxygen in place of toxic oxidizing agents.

We have therefore initiated a study of new bis(benzimidazole)-based diamide ligands, where the bound copper (II) is in a distorted five/six-coordinate environment with anodic $E_{1/2}$ values,⁹ and here we review this study. A series of new tetradentate diamide ligands N,N'-bis(2-benzimidazolylmethyl) hexanediamide (GBHA), N,N'-bis(N-octyl-2-benzimidazolylmethyl) hexanediamide (O-GBHA) and N,N'*bis*(2-benzimidazolylmethyl) 2,2'-oxy-diethaneamide have been synthesized and their Cu(II) complexes are reported. These have been found to carry out oxidation of various catechols and peroxidedependent facile oxidation of the aryl alcohols to aldehydes.

2. Synthesis

2.1 N,N'-bis(2-benzimidazolylmethyl) hexanediamide (GBHA)

This ligand was prepared following the procedure of Vagg et al^{10} . To a solution of adipic acid (1.67 g, 11.4 mmol) in pyridine (20 ml) was added a solution of glycine benzimidazole dihydrochloride (5.0 g,22.8 mmol) in pyridine (30 ml). The mixture was stirred gently for 10 min during which a white precipitate appeared. The reaction mixture was then heated slowly on a water bath at a temperature of 40°C, and to it was added triphenyl phosphite (7.08 ml, 22.8 mmol) dropwise over a period of 15 min. The reaction mixture was simultaneously stirred. After addition of P(OPh₃) was complete and the initially formed white precipitate dissolved, the temperature of the reaction mixture was slowly raised to 75°C and the clear solution was stirred for 1 h. A white solid resulted which was filtered off, washed with chloroform, and neutralized. The white crystalline product so obtained was recrystallised with a EtOH– H_2O (1:2) mixture and analysed for composition $C_{22}H_{24}N_6O_2$. H_2O .

Analysis: Found (calc): C 61.95 (62.16); H 6.14 (6.16); N 19.73 (19.90). m.p. 276°C. Yield: 1.9 g (41.1%) ¹H-NMR (d_6 -DMSO): δ (ppm) = 1.65 (quin, 4H), 2.26 (t, 4H), 4.54 (d, 4H), 7.15–7.49 (m, 8H), 8.44 (t, 2H_{amide NH}), 12.07 (s, br, 2H_{benzim NH}), IR (KBr Pallets): 3296 cm⁻¹ ($v_{N-H amide}$), 3185 cm⁻¹ ($v_{N-H benzim}$), 1635 cm⁻¹ ($v_{C=0 amide 1}$), 1539 cm⁻¹ ($v_{C-N amide II}$), 1448 cm⁻¹ ($v_{C=N-C=C benzim}$). A_{max}, nm (log ε) (MeOH): 243 (3.67), 272 (3.68), 279 (3.68). 2.2 *N*,*N'*-bis(*n*-octyl-2-benzimidazolylmethyl) hexanediamide (*O*-GBHA)

The N-octyl derivative of GBHA was synthesized by modifying the procedure of Reed *et al.*¹¹

2.3 N,N'-bis(2-methylbenzimidazolyl) 2,2'-oxydiethanamide (GBOA)

This ligand was prepared in a similar manner as GBHA except diglycollic acid was used instead of adipic acid. The reddish brown solution obtained was cooled down to room temperature, washed with saturated solution of sodium bicarbonate till all effervescence ceased and then washed twice with distilled water. Immediately a yellowish white solid appeared which was first washed with water and then with acetone to give white flaky solid, which was recrystallised using EtOH–H₂O (1:2) and analysed for composition $C_{20}H_{20}N_6O_3$ ·2H₂O.

Analysis: Found (calc): C 56·06 (56·07), H 5·59 (5·61), N 19·78 (19·63)%. m.p.: 125°C, yield: 2·6 g (59·2%), ¹H-NMR (DMSO- d_6): δ (ppm) = 4·11 (*s*, 4H), 4·57 (*d*, 4H), 7·11–7·14 (*q*, 4H), 7·47–7·50 (*q*, 4H), 8·73 (*t*, 2H_{amide NH}), IR bands (cm⁻¹, KBr pallets): 3203 ($v_{\text{N-H amide}}$), 3158 ($v_{\text{N-H benzim}}$), 1657 ($v_{\text{C=O amide II}}$), 1543 ($v_{\text{C-N amide II}}$), 1446 ($v_{\text{C=N-C=C benzim}}$), 1134 ($v_{\text{C-O-C}}$). λ_{max} nm (log ε) (MeOH): 279 (4·12), 272 (4·18), 247 (4·07), λ_{max} nm (log ε) (2 : 8 DMSO: CH₃CN): 281 (4·39), 277 (4·41).

3. Synthesis of complexes

3.1 [CuCl (GBHA)] Cl

To a methanolic solution of $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$ (0.5 mmol) was added a methanolic solution of the ligand (0.5 mmol). The resulting parrot-green solution was stirred for 1 h, after which the volume was reduced on a warm water bath. The parrot-green product obtained was washed with a small amount of methanol and air-dried. It was then redissolved in excess warm methanol, and the clear solution was left undisturbed for weeks to give beautiful green needles of the complex, suitable for X-ray diffraction analysis. Crystals were analysed for the composition $\text{CuC}_{22}\text{H}_{24}\text{N}_6\text{O}_6$ $\text{Cl}_2 \cdot \text{H}_2\text{O} \cdot \text{CH}_3\text{OH}$.

Analysis: Found (calc): C 46·72 (46·89); H 5·05 (5·10); N 14·40 (14·27)%. λ_{max} , nm (log ε) (MeOH): 758 (2·05), 360 (*sh*), 277 (4·16), 271 (4·18). $\mu_{eff}(\mu_B) =$ 1·90. $E_{1/2}$ (*V*) = +0·584 vs NHE. IR (cm⁻¹, KBr);

3395 ($v_{\text{N-H amide}}$), 3209 ($v_{\text{N-H benzim}}$), 1604 (amide 1), 1560 (amide II), 1452 ($v_{\text{C=N-C=C}}$).

3.2 $[Cu(NO_3)(GBHA)](NO_3)$

This complex was prepared in the same manner taking $Cu(NO_3)_2 \cdot 3H_2O$ in place of $CuCl_2 \cdot H_2O$. The green product was washed with methanol, dried in air, and analysed for the composition $CuC_{22}H_{24}N_8O_8 \cdot$ $0.5H_2O \cdot 0.5 CH_3OH$.

Analysis: Found (calc): C 43·81 (43·79); H 4·33 (4·38); N 18·30 (18·17)%. λ_{max} , nm (log ε) (MeOH): 782 (1·94), 347 (*sh*), 277 (4·28), 270 (4·30). μ_{eff} (μ_{B}) = 2·04. $E_{1/2}$ (*V*) = +0·599 vs NHE. IR (cm⁻¹, KBr); 3230 ($v_{N-H \text{ amide}}$), 3097 ($v_{N-H \text{ benzim}}$), 1602 (amide 1), 1501 (amide II), 1445 ($v_{C=N-C=C}$), 1384 and 821 ($v_{O-N-O \text{ sym and antisym}}$).

3.3 [*Cu*(*NO*₃)(*GBHA*)]

To a methanolic solution (10 ml) of GBHA (0.5 mmol) was added a methanolic solution (5 ml) of Cu $(NO_3)_2 \cdot 3H_2O$ (0.5 mmol). The resulting parrot-green solution was stirred for 1 h in a septum-sealed threeneck flask. A solution of quinol (0.5 mmol) was prepared in methanol (10 ml). High purity dry nitrogen was then bubbled through both the solutions for 30 min. The quinol solution was then transferred under nitrogen, via a double-end needle, to the copper (II) complex solution. An immediate colour change from green to yellow was observed. Stirring for 10– 15 min resulted in a white solid that was filtered off, washed several times with methanol, and dried *in vacuo*.

Analysis: Found (calc) for $\text{CuC}_{22}\text{H}_{24}\text{N}_7\text{O}_5$ · 0·7CH₃OH. C 49·63 (49·32); H 4·77 (4·88); N 17·61 (17·70)%. $E_{1/2}$ (V) = +0·622 vs NHE. IR (cm⁻¹, KBr); 3275 ($v_{\text{N-H amide}}$), 3195 ($v_{\text{N-H benzim}}$), 1653 (amide 1), 1538 (amide II), 1459 ($v_{\text{C=N-C=C}}$), 1384 and 821 ($v_{\text{O-N-Osym and antisym}}$). ¹H-NMR of freshly precipitated [Cu(NO₃)–(GBHA)] (slightly wet with methanol) in d_6 -DMSO: δ (ppm) = 1·54 (*quin*, *br*, 4H), 2·21 (*t*, *br*, 4H), 4·73 (*d*, *br*, 4H), 7·30–7·86 (*m*, *br*, 8H), 8·83 (*t*, *br*, 2H_{amide NH}).

$3.4 [CuCl_2(GBOA)]$

This was prepared in the same manner as the chloride complex of GBHA. The olive-green product was filtered, washed with small amounts of ethanol and air-dried. The product was recrystallized from hot ethanol and analysed for the composition $CuC_{20}H_{20}N_6O_3Cl_2\cdot 2H_2O$ Yield: 200 mg (70%).

Analysis: Found (calc): C 42·35 (42·67), H 4·24 (4·27), N 14·85 (14·93)%, λ_{max} nm (log ε) (2:8 DMSO: CH₃CN): 780 (1·93), 378 (2·77), 279 (4·10), 272 (4·14), μ_{eff} (BM): 1·82; $E_{1/2}$ (V): +0·574 V vs NHE. IR bands (cm⁻¹, KBr pallets): 3258 ($v_{N-H amide}$), 1654 ($v_{amide I}$), 1542 ($v_{amide II}$), 1457 ($v_{C=N-C=C}$). Crystals suitable for X-ray diffraction analysis were grown by dissolving the green complex in warm ethanol, layering it with acetone and leaving the solution for a few weeks to obtain green needles of the complex.

3.5 $[Cu^{1}(NO_{3})(GBOA)]$

This was prepared in the same manner as $[Cu^{1} (NO_{3})GBHA]$. The product was analysed for the composition $CuC_{20}H_{20}N_{7}O_{6}.0.5$ H₂O. Yield: 170 mg (65%).

Analysis: Found (calc): C 45.55 (45.58), H 4.00 (3.99), N 18.60 (18.61)%, $E_{1/2}$ (*V*): +0.395 V vs NHE. λ_{max} nm (log ε) (DMSO): 279 (4.12), 272 (4.17). IR bands (cm⁻¹, KBr pallets): 3238 ($v_{\text{N-H amide}}$), 3111 ($v_{\text{N-H benzim}}$), 1649 ($v_{\text{amide II}}$), 1541 ($v_{\text{amide II}}$), 1458 ($v_{\text{C=N-C=C}}$), 1384 and 821 ($v_{\text{O-N-O sym and antisym}}$) ¹H-NMR of freshly precipitated [Cu¹ (NO₃) (GBOA)] (slightly wet with methanol) (DMSO- d_6): δ (ppm) = 4.11 (*s*, *br*, 4H), 4.37 (*s*, *br*, 4H), 7.34 (*s*, *br*, 4H), 7.84 (*s*, *br*, 4H), 9.07 (*s*, 2H_{amide NH}).

The same product was obtained when the reducing agent was altered from quinol to ascorbic acid and to dihydroxyacetophenone. Similarity of the products was confirmed by analytical data and ¹H-NMR spectra.

3.6 $[Cu(O-GBHA)(NO_3)](NO_3)$

This was prepared by a procedure similar to that used for the above complex, by taking $Cu(NO_3)_2$. $3H_2O$ in place of $CuCl_2 \cdot 2H_2O$. The bright-green complex obtained was air-dried. The complex was redissolved in $CHCl_3$: MeOH (5:1, v/v) and was layered with petroleum ether. The clear solution was left undisturbed for weeks to give green crystals, suitable for X-ray diffraction work. The complex was analysed for the composition $CuC_{38}H_{56}N_6O_2(NO_3)_2$.

Analysis: Found (calc): C 56·24 (55·91), H 6·91 (6·86), N 13·86 (13·73)%, $\lambda_{max} nm (log \varepsilon) 275 (4·12)$, 283 (4·06), 740 (1·95).

4. Structural details

4.1 $[CuCl(GBHA)]Cl \cdot H_2O \cdot CH_3OH$

The structure is shown in figures 1 and 2. The crystals are stable only in mother liquor and tend to become opaque when taken out. Crystals have the space group C2/c of a monoclinic system. The coordinate environment around the copper atom is pentacoordinate consisting of two benzimidazolyl nitrogen atoms, two amide carbonyl oxygen atoms, and a chloride ion. The structure has the appearance of a trigonal bipyramid in which equatorial positions are occupied by a Cl atom and amide carbonyl oxygen atoms O(1) and O(2), and the axial positions by benzimidazole imine nitrogen atoms N(1A) and N(1B). Cu-N bond distances of 1.967 Å (Cu-N(1A)) and 1.977 Å (Cu–N(1B)) are in the range found for similar benzimidazole and imidazole ligated compounds¹². Cu-O bond distances of 2.111 Å (Cu-O(1)) and 2.166 Å (Cu–O(2)) are considerably longer than those reported in the literature for amide carbonyl coordination.13



Figure 1. ORTEP projection of [Cu(GBHA)Cl] Cl.H₂O CH₃OH showing the atomic numbering scheme.



Figure 2. Stereo drawing of the unit cell contents of $[Cu(GBHA)Cl]-Cl\cdot H_2O-CH_3OH$.

In pentacoordinated systems, the actual geometry of the complex can be described by a structural index parameter τ such that $\tau = \beta - \alpha/60^\circ$, where β and α are the two largest angles ($\beta > \alpha$). According to this model, out of the equatorial ligands (A, D, E), A is chosen such that the Cu-A bond length is longer than Cu-D/E and should not be any one of the four donor atoms forming the two largest angles. Thus, the geometric parameter τ is applicable to pentacoordinated structures as an index of the degree of trigonality, within the structural continuum between trigonal bipyramidal (TBP) ($\beta - \alpha = 60^{\circ}, \tau = 1$) and square pyramidal ($\beta - \alpha = 0^{\circ}$, $\tau = 0$). The complex has a $\tau = 0.591$, indicating a distortion from the perfect TBP towards distorted TBP. This complex does not fit the above described model fully, since the Cu–(2) (Cu-A) bond length of 2.166 Å is shorter than the Cu–Cl (1) (Cu–D/E) bond length of 2.3082 Å. This observed Cu-Cl(1) is in the range similar to that observed for similar pentacoordinated Cu(II) compounds¹⁴. This coordinated Cl atom is also hydrogenbonded to the benzimidazole NH of the neighbouring complex molecule. Equatorial angle deviates largely from the expected value of 120°, the largest deviation being 24.79°, whereas N(1A)-Cu-N(1B) is 176.89(13)°. The sum of equitorial angles at Cu is 359.95°, showing that the metal ion is essentially in the mean plane of the equatorial donors. The crystal consists of layers of [CuCl (GBHA)]Cl·H₂O·CH₃OH stacked over one another, held together by NHamide...Owater and NHamide... O_{MeOH} interactions, whereas molecules within the layers are H-bonded to each other via NH_{benim}...Cl bonds.

4.2 $[CuCl_2(GBOA)]$

The structure is shown in figure 3. The Cu(II) ion is surrounded by two nitrogen atoms (N2 and N6), two



Figure 3. ORTEP projection of [Cu(GBOA)Cl₂] showing the atomic numbering scheme.

chlorine atoms (Cl1 and Cl2) and one oxygen atom occupying the vertex of a square-based pyramid. Two *trans*-bonded chlorine atoms (Cl1 and Cl2) and nitrogens (N1 and N6) form the equatorial plane. The apical position is occupied by the oxygen atom (O3) of an amide carbonyl group. It is a one-dimensional polymer structure with the bonding scheme: ...Cu...ligand...Cu...ligand – such that the two benzimidazole units coordinated to a given Cu(II) ion are from different ends of two different molecules of the ligand.

The equatorial Cu–N bond distances of 1.968 Å (Cu–N (2)) and 1.985 Å (Cu–N (6)) are in the range found for similar benzimidazole and imidazole-ligated compounds.^{12,15} These are only slightly longer than those of GBHA ligated copper complex CuCl₂GBHA. Cu–Cl bond distances of 2.362 Å (Cu–Cl (1)) and 2.336 Å (Cu–Cl (2)) are in the range reported for compounds containing equatorially bound chlorine atoms.

The axial Cu-O3 bond length of 2.390 Å is considerably longer than those reported in the literature for Cu(II) complexes having axial oxygen coordination. The complex has a τ value of 0.167 ($\tau = \beta - \alpha/60^\circ$) where $\alpha = N2-Cu-N6 = 164.5^{\circ}$ and $\beta = Cl2-Cu-Cl1 =$ 174.52°, indicating square pyramidal geometry with a small trigonal component. The equatorial angles deviate slightly from the expected value of 90°, the maximum deviation being 2.6° . Two trans chlorine atoms are at an angle of 174.52° and the trans nitrogen atoms are not completely linear, the N2-Cu–N6 bond angle being 164.5°. The bonding of N2– Cu-N6 axis indicates that the two benzimidazole rings are not on the same plane. The sum of equatorial bond angles is 358.8° indicating that Cu(II) atoms mainly reside in the square plane with only slight displacement above the plane towards apical oxygen. The apical Cu-O bond is also not perpendicular to the square plane, the angles O3-Cu-N2, O3-Cu-N6 and O3-Cu-Cl1 being about 97°. On an average, the lengthening of Cu-N and Cu-O bonds in CuCl₂GBOA can be correlated with the reduced flexibility of the ligand due to shortening of the chain compared to GBHA, as a result of which the closer approach of donor to metal ion is slightly hindered. This also results in ligand-bridged mode structure of these complexes. The expanded angles indicate the displacement of equatorial atoms away from the apical oxygen. Crystals of the Cu(I) complex could not be grown despite our best efforts.

4.3 $[Cu(NO_3)(O-GBHA)](NO_3)$

The structure is shown in figure 4. The copper centre in the complex has tetragonally distorted octahedral environment. The six-coordination centre comprises two imine nitrogens of the two benzimidazole rings, two amide oxygens of the ligand and two oxygens of one of the nitrate anions. The carbonyl oxygen atom O2, nitrate oxygen atom O3, and the two benzimidazole imine nitrogen atoms N4 and N5 occupy the equatorial positions of the plane while the axial positions are occupied by another amide carbonyl oxygen atom O1 and nitrate oxygen atom O5 in distorted octahedral geometry. Bond lengths show that the nitrate group behaves as a highly unsymmetrical bidentate chelating ligand, with Cu-O5 bond length 2.46 Å much longer than Cu-O3 bond length 2.074 Å. The bond angle O3–Cu–O5 $55.1(8)^{\circ}$ is in the range found for weakly coordinating bidentate ligands. The Cu-N bond distances of 1.983 Å (Cu-N4) and 1.973 Å (Cu–N5) are found to be similar to those reported typically with imidazole/benzimidazole N-atoms in the equatorial plane.^{12,15} Thus, the short distances in the present Cu(II) complex probably reflect that the overall positive charge on the complex is not fully compensated by the unsymmetrical bidentate nitrate ligand as compared to those complexes with good coordinating anions. The Cu–O bond distances of 2.020 Å (Cu–O2) and 2.212 Å (Cu–O1) are considerably longer than those reported in the literature for amide carbonyl coordination. The bond angles are quite close to the values expected for an octahedral structure except for O1-Cu-O5 156.5(7)° and O3–Cu–O5 $55 \cdot 1(8)^{\circ}$ which are on the smaller side. The distortion may be due to the combined influence of the Jahn-Teller effect and the fixed bite size of the NO_3^- ion. The atoms O1, O2, O3, O5 and O2, O3, N4, N5 lie in approximate planes with minimum deviation of O5 being 0.05 Å for O5. These three planes



Figure 4. ORTEP view of [Cu(O-GBHA)(NO₃)](NO₃) showing the rotation about the C3–C4–C5–C6 bond.

are almost perpendicular to each other; the dihedral angles between the first and second and first and third are 85.5 (3) and $86.7(3)^{\circ}$ respectively, whereas between the first and third is $76.9(4)^{\circ}$. Two benzimidazole rings are planar and have a dihedral angle of $29.4(3)^{\circ}$ with respect to each other. Torsional angles (Cu-N4-C10-C15, 8°; Cu-N5-C17-C22, 6°(3)' also N4-C10-C15-C44, 179(2)°; N5-C17-C22-C21, $-178(2)^{\circ}$ show that the two benzimidazole rings are projected in opposite directions to each other. The conformation of the chain joining the two benzimidazole rings may be defined as gaagagaag. A kink at C3-C4-C5-C6 gives a spiral turn to the ligand, and it wraps around the metal ion in the form of a half helix. The packing of the molecule shows an extensive hydrogen-bonding network. It involves intramolecular hydrogen bonds between amide nitrogen N2 and N6 and N8 of the free nitrate group. The second amide nitrogen N1 gives two strong intermolecular Hbonds with O7 and N8 of the symmetry related uncoordinated nitrate group. Apart from these, there are a number of weak C-H...O and C-H...N Hbonding interactions involving both the nitrate ions and various methylene and phenylene carbons.

4.4 $[Cu(NO_3)(GBHA)](NO_3) \cdot CH_3OH$

The crystal structure of the compound is shown in figure 5. It consists of discrete monomeric units composed of a cationic copper complex and a nitrate anion. The coordination geometry around the Cu(II)



Figure 5. ORTEP projection of [Cu(GBHA)(NO₃)] (NO₃)·CH₃OH showing atomic numbering scheme.

ion is distorted square pyramidal with benzimidazole imine nitrogen atoms, carbonyl oxygen atoms and nitrate oxygen atom occupying the vertices. Two trans bonded benzimidazole nitrogen atoms (N1 and N1'), an amide carbonyl oxygen atom (O2) and a nitrate oxygen atom (O1'), form the square plane while the pyramid is completed by second amide carbonyl oxygen (O2'). The Cu-N_{benzim} distances of 1.973 Å [Cu-N (1)] and 1.975 Å [Cu-N (2)] are in the range found for similar complexes. Cu-O distance of 1.992 Å for equatorially bound carbonyl oxygen is in the expected range while the Cu-O bond distance of 2.229 Å for axially bound carbonyl oxygen is considerably longer than those reported in literature.¹³ The nitrate ion coordinates to Cu(II) in monodentate fashion through O1'. N4–O1' bond length (1.267 Å) for nitrate is longer than the two N4-O1 and N4-O3 bonds of nitrate. The lengthening of this bond as compared to remaining two bonds is expected because of transfer of electron density from oxygen to Cu(II) and weakening of the N4-O1' bond. The Cu-O1' bond length of 2.061 Å is well close to the literature value of 2.000 Å. Bond angles of axial oxygen with equatorial atoms indicate a distortion of the equatorial plane away from axial oxygen. Thus O2, N1', O1' are displaced below the equatorial plane while N1 is displaced above the plane. Coordination of one amide oxygen at axial position indicates that two symmetrical halves of the ligand may be in different planes. The actual geometry of the five-coordinated complex is described by the geometric parameter τ which is defined as an index of degree of trigonality, within the structural continuum between triagonal bipyramidal ($\tau = 1$) and square pyramidal ($\tau = 0$) such that $(\tau = \beta - \alpha/60^\circ)$ where β and α are the two largest angles $(\beta > \alpha)$. τ value for this complex comes out to be 0.168 indicating the slight distortion of square pyramidal geometry.

5. ¹H NMR, electronic spectrosopy, and cyclic voltammetry

The electronic spectra of all the complexes were measured in methanol. UV bands at 272 and 279 nm are observed for the free ligand GBHA. These bands are characteristic of the benzimidazole group and arise from a $\pi \rightarrow \pi^*$ transition. All the complexes display a broad *d*–*d* band in the region 740–790 nm characteristic of tetragonal geometry. A shoulder due to a LMCT (imidazole $\pi_1 \text{ Cu}^{2+}$) band in the region 340–390 nm is also observed. The UV band in the

(a)

complexes corresponding to a $\pi \rightarrow \pi^*$ transition shows enhanced absorption as indicated by their extinction coefficients.

The ¹H-NMR spectrum of the free ligand GBHA (figure 6a) shows signals for aliphatic and aromatic protons with theoretically predicted splittings. A symmetrical multiplet in the range 7.15-7.49 ppm arises due to benzimidazole ring protons characteristic of an AA"BB" pattern. A triplet at 8.44 ppm arises due to amide NH protons (coupled with adjacent CH₂ protons \bigcirc) (J = 5.5 Hz). Benzimidazole NH protons give a broad signal at 12.07 ppm. In the Cu(I) complex the aromatic ligand proton resonances are observed in two groups with a 1:3 proton ratio (figure 6b), unlike in GBHA where both the signals have the same proton integration. In the Cu(I) complex, the three aromatic protons (H₅, H₆, H₇) remain at the chemical shift close to that observed in the free ligand GBHA, while proton H₄ is shifted downfield, being nearest to the imidazole N atom bound to Cu(I). The signals are slightly broadened possibly due to the presence of trace Cu(II) ions. In copper (I) complex of [CuCl (GBOA)] (figure 7), slightly shifted and broadened signals were observed for all ligand protons with expected integration. ¹H signals here are downfield shifted compared to free ligand protons as they are in the vicinity of a Lewis acid Cu^+ and electronegative oxygen atoms of coordinated nitrate group which exert a deshielding effect through space electronic interaction.¹⁶ Broadening indicates that the protons are in a slightly paramagnetic surroundings which may be due to the presence of trace Cu(II) ions arising from a slight decomposition of Cu(I) complex.

All the GBHA-copper (II) complexes display a quasi reversible redox wave (figure 8) due to the copper(II)/Cu(I) process. Anodic shifts in $E_{1/2}$ values indicate the retention of the anion in the coordination sphere of Cu(II). The $E_{1/2}$ values vary anodically in the order $Cl^- < NO_3^- < SCN^-$. This indicates that bound thiocyanate destabilizes the Cu(II) state while bound chloride stabilizes it. The $E_{1/2}$ value for the copper (I) complex $[Cu^{1}(NO_{3}) (GBHA)]$ was found to be +0.622 V vs NHE. No redox wave associated with bound quinone or quinol was found in its cyclic voltammogram. When calculated against NHE, the $E_{1/2}$ values of the present series of Cu(II) complexes turn out to be quite positive as compared to those of other Cu(II) complexes with benzimidazole-based ligands (+0.00 to +0.446 V) and some amide based complexes. Binding of amide N, preferably in depro-

 r_{ppm} r_{g} $r_$

8.0

000

6.0

4.0

2.0

do

ıż n

iò.o

and (**b**) Cu(GBHA) NO₃ in d_6 -DMSO.

Figure 7. ¹H NMR spectra (in d_6 -DMSO) of (**a**) GBOA, (**b**) [Cu(GBOA)NO₃].





Figure 8. Cyclic voltammograms of (a) $Cu(GBHA)Cl_2$, (b) $Cu(GBHA)(NO_3)_2$, (c) $Cu(GBHA)(SCN)_2$ and (d) $Cu(GBHA)NO_3$ in a 2:8 DMSO-CH₃CN solution at a scan rate of 100 Mv/s.

tonated form, is known to stabilize copper in higher oxidation states (+2, +3), thus cathodically shifting the reduction potential values. In contrast the binding of an amide carbonyl oxygen apparently has a destabilizing effect on Cu(II), leading to relatively a high anodic redox potential for this series of complexes.

For the GBOA-copper (II) complexes, a quasi reversible redox wave due to the copper (II)/Cu(I) redox couple in 2:8 DMSO:CH₃CN mixed solvent system is observed. $E_{1/2}$ value of the complex [CuCl₂ (GBOA)] (+0.574 V) is in the same range as for GBHA complex. Cyclic voltammograms of copper complexes of O-GBHA were recorded in dichloromethane solution containing 0.1 M TBAP as supporting electrolyte at a glassy carbon working electrode. Ferrocene was used as internal standard, and potentials are referred to the ferrocenium/ferrocene (Fc⁺/Fc) couple. The cyclic voltammograms display a quasi-reversible wave at a fairly anodic potentials. This is attributed to the change of Cu(II) to Cu(I).

6. Electron exchange reaction between [Cu(NO₃)₂ (GBHA)] and *o/p*-dihydroxybenzenes

The reduction of $[Cu(NO_3)_2 (GBHA)]$ by quinol was studied spectrophotometrically under anaerobic conditions. A 7.8 mM [Cu(NO₃)₂ (GBHA)] solution was prepared in a 1:9 DMSO: CH₃CN solution. Dry nitrogen was bubbled through this septum-sealed solution for 30 min. A spectrum of the above complex in this solvent system displays a broad band centred at 747 nm ($\varepsilon = 154 \text{ M}^{-1} \text{ cm}^{-1}$). A 35.5 mM solution of quinol was prepared in a 1:9 DMSO: CH₃CN solution and dry nitrogen was bubbled through this septumsealed solution. A 0.04 ml sample of the above quinol solution was added to 2.0 ml of the Cu(II) complex solution in a septum-sealed quartz cuvette, with the help of an airtight syringe. The reduction of Cu(II) to Cu (I) was followed by observing the decrease in the optical density of the band centred at 747 nm. The same volume of quinol was added periodically, and the decrease in band intensity was monitored till 747 nm band almost vanished (figure 9a). At this point the Cu: quinol stoichiometry was 1:0.75. The colour of the solution faded to light yellow, indicating the disappearance of the Cu(II) species in the solution. The above yellow solution could be reoxidised by atmospheric oxygen to give a green solution, restoring the wavelength maximum at 747 nm. The same experiment was repeated with simple $Cu(NO_3)_2$ in the absence of the ligand GBHA. No fading of the light blue colour to yellow was observed. Instead the colour changed to green.

7. Catechol oxidase studies

A mixture of 2.0 ml of catechol solution (30 mM) in methanol (saturated with dry nitrogen) and 1.0 ml of copper complex solution (~3 mM) in methanol (saturated with dry dioxygen) was placed in a 1cm path length optical cell in a spectrometer. The final concentration of reaction mixture is catechol (20 mM) and complex (1 mM). The formation of o-benzouinone was followed by observing the increase of characteristic quinone absorption band in the range 390-410 nm (figure 9c). Blank run for catechol was also performed in the same manner by adding 1.0 mlof oxygen saturated methanol to 2.0 ml of catechol solution (30 mM) in nitrogen saturated methanol. Similarly blank run for complex was also performed. The same experiment was repeated using 4methyl-catechol (4M Cat), 3,5-ditertiary-butylcatechol, 3,4-dihydroxybenzoic acid and dopamine(Dop). For



Figure 9. (a) Vis spectra of a 7·8 mM solution of Cu(GBHA)(NO₃)₂ in a 2 : 8 DMSO–CH₃CN solution (curve 0) and Cu(GBHA)(NO₃)₂ in the presence of quinol (curve 1–8). (b) Visible spectra of a Cu(GBHA)(NO₃)₂ solution (1·1 mM, MeOH) (curve 0) and Cu(GBHA)(NO₃)₂ in the presence of catechol (curves 1 and 2). (c) UV-Vis spectra of catechol solution (20 mM, N₂ saturated methanol) [curve 0], new band generated in the presence of [Cu(GBOA)Cl₂] solution (1 mM, O₂ saturated methanol). Curve 2 – at 1 min, curve 3 – at 40 min. Curve 1 – charge transfer band of [Cu(GBOA) Cl₂] in blank experiment. (d) UV-Vis spectra of 4-methylcatechol solution (20 mM, N₂ saturated methanol) [curve 0], new band generated in the presence of [Cu(GBOA)Cl₂] solution (1 mM, O₂ saturated methanol). Curve 2, 2 min, curve 3, 66 min. Curve 4, 110 min, Curve 1, charge transfer band of [Cu(GBOA)Cl₂] observed in blank experiment.

each set of observation, a curve of concentration of o-benzoquinone formed (calculated by using corresponding ε values) versus time was plotted and initial rates were calculated by drawing a tangent to curve at t = 0 and finding its slope. After this initial fast phase, the average rate of reaction was also calculated.

7.1 *Kinetic studies of the reaction of* [*CuCl*₂*GBOA*] *with* 3,5-*ditertiarybutyl catechol*

The experiment described above was repeated with DTBC in two sets: (i) In the reaction mixture the concentration of catalyst [CuCl₂ (GBOA)] was varied

(0.5, 1.0.1, 2.0 mM) and concentration of DTBC was kept first at 20 mM. (ii) Concentration of DTBC was varied (10, 20, 30, 40 mM) and concentration of catalyst was kept fixed at 1 mM.

For each set of data initial rates were calculated and graphs of rate versus concentration of catechol were plotted.

For catechol, 4-methyl catechol and DTBC, the quinone production exceeds beyond the stoichiometric amount (table 1) while for dopamine, the *o*-benzouinone formed decayed very rapidly probably undergoing further reaction. With 3,4-DHB, no quinone generation could be observed, however the absorption in the region 390–410 nm increased continuously

Substrate	λ _{max} (nm) product <i>o</i> -enzoquinone	Initial rate (M min ⁻¹)	Average rate (M min ⁻¹)	Concentration of product formed at 35 min (mM)	Minimum conver- sion (mM) before decomposition of product	Catalyst turnover
Catechol	392	1.33×10^{-4}	2.6×10^{-5}	1.1114	1.4 (7%)	3
4-Methyl catechol	392	2.10×10^{-7}	9.3×10^{-4}	0.722	2.4 (12%)	5
3,5-Ditertiarybutylcatecho	ol 399	3.12×10^{-4}	2.25×10^{-5}	° 1.732	16.8 (84%)	34
Dopamine	380	Band decayed within 10 min				
2,4-Dihydroxybenzoic aci	d sh	Very week oulder decayed within 5 min				

Table 1. Rates of reaction observed with various substrates and [CuCl₂(GBOA)] as catalyst



Figure 10. Plot of *o*-benzoquinone (M), vs time for various catechols (20 mM); (a) 3,5-di-*t*-butylcatechol; (b) 4-methylcatechol; (c) catechol at [CuCl₂(GBOA)] =1 mM.

without displaying any well defined band indicating the occurrence of multiple reactions.

For catechol, DTBC, 4M cat, DOP and 3,4DHB, the concentration of corresponding quinone produced was calculated using $\varepsilon = 1585 \text{ M}^{-1} \text{ cm}^{-1}$, $1500^{17a} 2140$ and 2455 M^{-1} cm⁻¹,^{17b} respectively. For all these, quinone produced versus time curve follows biphasic behaviour (figure 10). The reaction occurs with a fast phase followed by a slower reaction, for both phases, the reaction rates have been calculated. Initial rates have been calculated by drawing a tangent to the curve at t = 0. For *o*-diphenol initial concentration is 20 mM and for [CuCl₂GBOA] initial concentration is 1 mM, the initial rates were found to be 1.33×10^{-4} , $2 \cdot 1 \times 10^{-4}$ and $3 \cdot 1 \times 10^{-4}$ M min⁻¹ for cat, 4 M Cat and DTBC respectively. For DTBC the conversion to DTBQ was 84.5% complete, i.e. turnover of 34 times was found within 27 h. During this time period, aerial oxidation of DTBC contributed to only 0.5% of the product DTBQ (confirmed by performing blank). Table 1 gives the conversion of various catechols to quinones at 30 min. The rates, though not comparable to the copper enzyme, tyrosinase, are found to be much higher than other synthetic complexes.^{17c} The activity is found to vary with the substituent on catechol as $HOOC < CH_2CH_2NH_2 < H < CH_3 < t$ -Bu. The order of reactivity shows that the o-diphenol where the substituent is the best electron-donating group gives the highest activity. Similar trends are found by other groups^{17b} also both for synthetic complexes and the enzyme tyrosinase. These results are consistent with the catecholase activity, which is dependent upon the formation of a Lewis acid-base bond between copper (II) and a o-diphenol oxygen atom. Based on this, absence of any quinine formation for 3,4-DHB could be understood in terms of a greater electron withdrawing effect of the carboxyl moiety, i.e. the COOH group in this particular catechol.

Kinetic studies have also been carried out using the substrate DTBC and the catalyst [CuCl₂(GBOA)]. For DTBC oxidation, the initial rate was found to depend linearly on the catalyst concentration while it is found to be dependent on the square root of the concentration of DTBC itself. It has been reported earlier that for a single mono-nuclear complex, the rate of DTBC oxidation depends linearly on the square of the copper (II) concentration, while our results indicate that it depends only on the first power of catalyst concentration. This implies that in the present series of complexes, the possibility of catechol bridging the two copper (II) centres is remote. This is quite likely due to the rigidity of bulky benzimidazole groups that may hinder the formation of such bridged species with catechol. Thus, the ratedetermining step involves only a single metal centre. Such an observation is not uncommon. This study mimics the activity of copper containing mettyrosinase and catechol oxidase enzymes.

8. Oxidation of 2,4,6-tri-*t*-butylphenol (TTBP)

Spectrophotometric analysis, of the reaction mixture consisting of TTBP and [Cu(NO₃)(GBHA)](NO₃) in the ratio 1:22 in MeOH saturated with dioxygen was carried out in the range 200–1100 nm (figure 11). A solution of pure TTBP in MeOH displays a weak band at 420 nm (log $\varepsilon = 0.477$) with a shoulder at 400 nm. On mixing TTBP with [Cu(NO₃)(GBHA)] (NO_3) an instant growth in bands at 400 and 420 nm is observed along with a new band at 318 nm, observed in a dilute reaction mixture. The generation of new bands at 318 and 400 nm indicated the formation of new products in the reaction mixture. The character of the original 400 nm shoulder also changes which starts to acquire the shape of a proper band with time. The band at 420 nm gradually loses its wellresolved shape to a shoulder with a shift in λ_{max} to 416 nm. The d-d band of [Cu(NO₃) (GBHA)](NO₃) (780 nm, $\log \varepsilon = 1.92$) in methanol shows an initial increase in intensity soon followed by a slight drop accompanied by a shift in λ_{max} to 770 nm (figure 12). This indicates the formation of active Cu(II) species.



Figure 11. UV-Vis spectra of (a) O_2 saturated methanolic solution (3.2 mM) of [Cu (NO₃)(GBHA)](NO₃) (b) O_2 saturated methanolic solution (65 mM) of TTBP (c)–(e) reaction mixture at 1, 15 and 30 min respectively after mixing of reactants. Inset. Diluted reaction mixture showing the band at 318 nm.



Figure 12. Visible spectra (d-d band) of (a) [Cu(NO₃) (GBHA)] (NO₃) (3·2 mM) in methanol. Reaction mixture containing catalyst (3·2 mM) and TTBP (65 mM) in O₂ saturated methanol at (a) 1 (b) 15 (c) 30 (d) 45 (e) 70 (f) 90 and (g) 120 min.



Figure 13. X-band EPR spectra of (a) $[Cu(NO_3) (GBHA)]$ (NO₃) in methanol, (b) reaction mixture at 1 min (c) 30 min at microwave frequency 9.4 GHz and receiver gain 10^3 .

A new band is also generated at 664 nm, which grows in intensity for about 70 min after which it starts to decay (λ_{max} shifting to 657 nm). The yellow colour of the reaction mixture turns green within this time period. This new band generated was assigned to tri-t-butylphenoxyl radical.¹⁸ The radical generation was also confirmed by low temperature (120 K) EPR measurements of the reaction mixture (figure 13). Pure [Cu(NO₃) (GBHA)](NO₃) in MeOH at 120 K displays a four-line EPR signal in the g_{\parallel} region and an unresolved signal in the g_{\perp} region with $g_{\parallel} = 2.40$ and $A_{\parallel} = 123$ G, $g_{\perp} = 2.093$. As soon as TTBP is added to the Cu(II) complex solution, a new signal is observed at field strength H = 3366.5 Hz and g = 2.004, this indicated the presence of free radicals in the reaction mixture. The signal grows with time, with a simultaneous drop in Cu(II) complex signal intensity. Thus both spectrophotometric and EPR measurements confirm the involvement of tri-t-butylphenoxyl radical in TTBP oxidation reaction.

9. Isolation of reaction products

Both the major products (A) 2,6-di-*t*-butyl-1,4benzoquinone and (B) 2,4-di-*t*-butyl-1,6-benzoquinone were isolated by column chromatography. A has $R_f = 0.73$ and B has $R_f = 0.3$ in benzene. They further purified through preparative TLC in yields 22% (A) and 8.6% (B). IR spectra (in KBr) of both the products display two new bands in the range 1600–1700 cm⁻¹ confirming the presence of carbonyl group in them. Product A has strong bands at 1656 and 1598 cm⁻¹ indicating two different types of carbonyls in the molecule. Product B displays two bands at 1656 and 1620 cm⁻¹, again due to two different types of carbonyls. This observation supports the benzoquinone type of structures for A and B.

¹H-NMR spectrum of the product A in d_6 -DMSO displays signals at δ (ppm) 1·24 (s, 18 H_a), 6·51 (s, 2 H_b). This observation supports *p*-benzoquinone structure: ¹³C-NMR of A in d_6 -DMSO give signals at δ (ppm) 189·4 (C-4), 188·2 (C-1), 158·3 (C-2, C-6), 142 (C-3, C-5), 35·2 (C-7), 29·7 (C-8). In mass spectrum, the base peak at *m/e* 220 also supports the above structure. Other peaks observed are at *m/e* 191, 179, 165, 149 and 107.

¹H-NMR spectrum of the product B in d_6 -DMSO displays signals at δ (ppm) 1·19 (s, 9H_a), 1·22 (s, 9H_b), 6·14 (d, 1H_c), 6·97 (d, 1H_d). The two aromatic protons are meta coupled, coupling constant J being 2·1 Hz. The presence of two different types of *t*-butyl

groups supports *o*-benzoquinone structure: ¹³C-NMR of B in d_6 -DMSO give signals at δ (ppm) 180 (C-1, C-2), 162.5 (C-4), 148.9 (C-6), 133.4 (C-3), 124 (C-5), 35.7 (C-7), 35.0 (C-9), 29.0 (C-10), 27.5 (C-10). In mass spectrum, the base peak at *m/e* 220 is observed.

Product A shows three bands in UV-Vis region 450 nm (log $\varepsilon = 1.59$), 318 nm (log $\varepsilon = 2.76$), 255 nm (log $\varepsilon = 4.57$) while product B shows bands at 537 nm (log $\varepsilon = 1.68$), 400 nm (log $\varepsilon = 2.93$), 255 nm (log $\varepsilon = 3.65$). The shift of band maximum towards longer wavelengths in B as compared to A confirms extended conjugation in B.

10. Mechanism of TTBP oxidation

The catalyst [Cu(NO₃)(GBHA)](NO₃) has copper in +II oxidation state and is in a square-pyramidal coordination environment. The first step possibly involves the deprotonation of TTBP whereby the TTBP anion coordinates to Cu(II). The Cu(II) complex is capable of oxidizing TTBP to TTBP radical simultaneously producing the reduced catalyst, i.e. Cu(I) species. The appearance of a band at 664 nm and the appearance of a sharp EPR signal at g = 2.004 confirms the formation of the phenolate radical. The reduced catalyst Cu(I) is then reoxidised by molecular oxygen to give the superoxide radical (scheme 1).

The super oxide anion radical generated then attacks at the para position of rearranged TTBP radical to produce an intermediate hydroperoxy compound which rapidly eliminates a molecule of t-BuOH to give product A. In yet another competitive reaction,



Scheme 1. Catalytic generation of TTBP radical by [Cu(NO₃)(GBHA)] (NO₃).



Figure 14. Mechanism of TTBP oxidation.



Figure 15. Curve 1, d-d band spectrum of a reaction mixture (0.0308 mmol of CuCl(O-GBHA)]Cl, 0.616 mmol of 4-chlorobenzyl alcohol, 0.616 mmol of cumenyl hydroperoxide) at the time of mixing. Curves 2–4, d-d band spectra of a reaction mixture after every 10 min of heating curve 5, d-d band spectrum of a reaction mixture 1 h after the initiation of reaction.

the TTBP radical undergoes rearrangement to produce an unpaired electron at the ortho position which undergoes attack by TTBP radical in the same manner to give product B. The present study thus indicates that the complex [Cu(NO₃)(GBHA)](NO₃) is able to carry out oxidative hydroxylation with simultaneous dealkylation reaction in organic substrates and is an example of activation of molecular oxygen (figure 14).

11. Oxidation of alcohols

A 25 ml, two-necked flask was fitted with a reflux condenser and an oxygen/nitrogen inlet. Copper complex [CuCl(O-GBHA)]Cl (23.5 mg; 0.0308 mmol) and chlorobenzyl alcohol (87.83 mg; 0.616 mmol) in 10 ml CH₂Cl₂ were stirred at room temperature for 10 min under nitrogen. Cumenyl hydroperoxide (0.088 ml; 0.616 mmol) was added and stirred for 20 min. At this point the d-d band of the copper complex was monitored (figure 15). This was followed by raising the temperature of the reaction mixture to 40–45°C on a water bath. Stirring was continued for 1-2 h. The reaction was monitored by visible spectroscopy, and it was found that the d-d band intensity drops to 50% of its original value, within a 30 min period (figure 15), and subsequently remains constant for the remaining period of the reaction. No new band is generated during the reaction cycle in the region 300-1100 nm. The reaction mixture was further stirred for another 24 h at room temperature. TLC of the reaction mixture showed two major products that were visualized by spraying an ethanolic solution of 2,4-DNP. The reaction products were finally separated by column chromatography using 10% ethylacetate/hexane as eluent (one of them was identified as the corresponding aldehyde and the other as acetophenone). Products separated by column chromatography were quantitatively estimated using HPLC. A standard curve for each of the aldehydes was generated, and the concentration of the unknown aldehyde was obtained with reference to the standard curve. On this basis, the percentage conversion and turnover numbers are reported in table 2.

Identity of the aldehydes and acetophenone formed was confirmed by ¹H-NMR of their respective DNP derivatives and are in confirmation with those reported in the literature. No products were obtained if the solution was not warmed to $40-45^{\circ}$ C for 30 min, and the oxidation state of Cu(II) also does not change if the reaction mixture is not heated as above; this is confirmed by observing minimal change in *d*–*d* band

	Catalyst – [Cu(OctGBHA)Cl ₂]			
Substrate	ArCHO (% yield) (TON)	PhCOCH ₃ (% yield) (TON)		
Benzyl alcohol	38.96 [16]	27.8 [16]		
4-Chlorobenzyl alcohol	43.3 [18]	42.3 [23]		
4-Ethylbenzyl alcohol	51.93 [18]	47.1 [26]		
4-Methylbenzyl alcohol	64.94 [27]	27.8 [16]		
4-Nitrobenzyl alcohol	32.5 [13]	27.8 [16]		
1-Phenylethanol	_	65.70 [32]		

Table 2. Percentage yield of products obtained on oxidation of various alcohols using [CuCl(O-GBHA)]Cl as catalyst.

TON = turnover number

intensity for such room temperature solutions, for a period of several hours. Similar results were obtained using $[Cu(NO_3)(O-GBHA)](NO_3)$ as catalyst.

It is well established that in the absence of protic solvent, the hydroperoxides are likely to cleave by homolysis reaction.¹⁹ Further it has been reported that cumenyl hydroperoxide generates a cumylperoxyl radical (R_3COO^{\bullet}) with metal ion complexes with concomitant change in oxidation state of the metal ion.²⁰ It is also established that cumylperoxyl radicals decompose by a mechanism which finally generates acetophenone as one of the products.²¹ The above two reaction sequences are presented as:

$$\begin{split} &R_{3}\text{COOH} + \text{Cu(II)} \rightarrow \text{R}_{3}\text{COO}^{\bullet} + \text{Cu(I)} + \text{H}^{+}, \\ &R_{3}\text{COOH} + \text{Cu(I)} \rightarrow [\text{R}_{3}\text{CO}...\text{Cu(II)}] + \text{OH}^{-}, \\ &R_{3}\text{COO}^{\bullet} \rightarrow \text{R}_{3}\text{CO}^{\bullet} + 1/2\text{O}_{2}, \\ &R_{3}\text{CO}^{\bullet} \rightarrow \text{PhCOCH}_{3} + \text{R}^{\bullet}. \end{split}$$

It is assumed that the above reaction sequences are dominantly active in the present catalytic oxidation of alcohol. A spectral monitoring of the course of catalytic reaction shows a drop in d-d band intensity to almost half of its original value after thermally activating the reaction for nearly 30 min. This may imply the reduction of copper (II) to copper (I). However, there appears to be some structural change which takes place during the activation; the d-dband shifts by almost 30 nm (λ_{max} : 712 nm) to shorter wavelengths and a weak shoulder around 880nm is also lost, when [CuCl(O-GBHA)]Cl complex is used as a catalyst. This possibly implies a loss of the axially bound atom in the complex leading to formation of an "active copper species".²²

In an attempt to identify the Cu(II) species that may be active in the peroxide-dependent oxidation of aryl alcohols, a sample of Cu(II) species (which is

formed after the reaction is thermally activated for 30 min in the presence of substrate and cumenyl hydroperoxide) was isolated and EPR of this sample was taken in CH₂Cl₂ at liquid nitrogen temperature. For comparison, the EPR of the original complex [CuCl(O-GBHA)]Cl was utilized in this reaction, as a catalyst was also taken in the same solvent and at liquid nitrogen temperature. A comparison of the EPR spectra of the two species indicates that there is slight variation in their respective g-values; however, they follow the order $g_{\parallel} > g_{\perp} > 2.0023$, implying a $d_{x^2-y^2}$ ground state. There are, however, changes in the hyperfine coupling constants ([CuCl(O-GBHA)]Cl) prior to reaction, $g_{\parallel} = 2.40$, $g_{\perp} = 2.12$, and $A_{\parallel} = 120$ G; ([CuCl(O-GBHA)]Cl) after the reaction, $g_{\parallel} = 2.34$, $g_{\perp} = 2.09$, and $A_{\parallel} = 135$ G). The $g_{\parallel}/10^{-4} A_{\parallel}$ index is 200 for the original complex while it is 173 for the active species. This implies that the active species is more planar relative to the starting complex.²³ This relative planarity may result due to the relieving of strain in the complex caused by the loss of the axially bound atom in the "active-copper species".

The role of molecular oxygen was also studied by allowing a slow stream of O_2 during the oxidation reaction catalysed by the [CuCl(O-GBHA)]Cl complex. The change observed in the d-d band of the catalyst is depicted in figure 16a. It is found that the activecopper species with a $\lambda_{max} \sim 712$ nm is again generated, but in this case there is no drop in the absorbance of the d-d band of the catalyst; rather, there is slight increase in the absorbance. This implies a higher extinction coefficient of the d-d band for the activecopper (II) species relative to the starting complex. Further, the added molecular oxygen competes strongly with step 2 of the proposed reaction cycle and keeps most of the copper in the oxidized state. This has direct impact on the yields of the two products, viz. aldehyde and acetophenone, that drop dramatically; the aldehyde yield decreases by 80% while the ace-



Figure 16. (a) Curve 1: d-d band spectrum of a reaction mixture (0.0308 mmol of [CuCl(O-GBHA)]Cl, 0.616 mmol of 4-chlorobenzyl alcohol, 0.616 mmol of cumenyl hydroperoxide) under O₂ at the time of mixing. Curve 2: d-d band spectrum of a reaction mixture 1 h after the initiation reaction. (b) Curve 1: d-d band spectrum of a reaction mixture (0.0308 mmol of [CuCl(O-GBHA)] Cl, 0.616 mmol of 4-chlorobenzyl alcohol, 0.616 mmol of cumenyl hydroperoxide, 0.308 mmol of [CuCl(O-GBHA)] Cl, 0.616 mmol of 4-chlorobenzyl alcohol, 0.616 mmol of cumenyl hydroperoxide, 0.308 mmol of TBHP) at the time of mixing. Curve 2: d-d band spectrum of a reaction mixture after 30 min of heating. Curve 3: d-d band spectrum of a reaction mixture 1 h after the initiation of reaction.



Figure 17. Proposed catalytic reaction cycle.

tophenone by 46%, indicating severe competition by oxygen in steps 2 and 3 of the reaction cycle.

The generation of cumylperoxyl radicals during the course of the present reaction is confirmed by the identification of acetophenone as one of the reaction products. Further, when a radical trap like 2,4,6-trit-butylphenol (0.308 mmol, TBHP) is included in the reaction mixture and the reaction is carried out as indicated under the oxidation of alcohols (with vigorous exclusion of oxygen), it is found that a new band at ~ 640 nm generate is after the reaction has been thermally activated for 30 min. This supports the formation of the stable 2,4,6-tri-*t*-butylphenoxy radical²² via the oxidation of TBHP by cumylperoxyl radicals (figure 16b).

The yield of the aldehyde and acetophenone obtained after workup of the reaction indicates a drop of 74% for the ketone (table 2). This implies that the presence of the radical trap severely inhibits step 3 relative to step 1 of the proposed reaction cycle. This further justifies the presence and the utilization of cumylperoxyl radical in the oxidation of 2,4,6-tri-*t*-butylphenol to the phenoxy radical.

The proposed catalytic reaction cycle is depicted in figure 17. This reactivity is reminiscent of the functioning of copper site in the enzyme galactose oxidase.

Thus the present series of *bis*-benzimidazole diamide Cu(II) complexes are capable of activating the peroxyl group and molecular oxygen.

Acknowledgement

We gratefully acknowledge financial support from the Department of Science and Technology, Govt. of India.

References

- 1. Solomon E I, Baldwin M J and Lowery M D 1992 Chem. Rev. 92 521
- 2. Klabunde T, Eicken C, Sacchettini J and Krebs B 1998 Nat. Struct. Biol. 5 1084
- (a) Klinman J P 1996 Chem. Rev. 96 2541; (b) Whittaker J W and Whittaker M M 1998 Pure Appl. Chem. 70 903; (c) Whittaker J W and Sigel A 1994 Met. Ions Biol. Syst. 30 315
- 4. Whittaker M M, De Vito, V L, Asher S A and Whittaker J W 1989 J. Biol. Chem. 264 7104
- (a) Ito N, Philips S E V, Stevens, C, Orgel Z B, Mc Pherson M J, Keen J N, Yadav K D S and Knowles P F 1991 *Nature (London)* **350** 87; (b) Ito N, Philips S E V, Yadav K D S and Knowles P F 1994 *J. Mol. Biol.* **238** 794
- Anson F C, Collins T J, Richmond T G, Santarsiero B D, Toth J E and Treco B G R T 1987 J. Am. Chem. Soc. 109 2974
- Koikawa M, Okawa H, Maeda Y and Kida S 1992 Inorg. Chim. Acta 1992 19475
- (a) Redmore S M, Rickard C E F, Webb S J and Wright L J 1997 Inorg. Chem. 36 4743; (b) Patra A K, Roy M and Mukherjee R 1999 J. Chem. Soc., Dalton Trans. 2461; (c) Kawamoto T, Hammes B S, Ostrander R, Rheingold A L and Borovik A S 1998 Inorg. Chem. 37 3424
- Gupta M, Mathur P and Butcher R J 2001 Inorg. Chem. 40 8(b); Gupta M, Das S K, Mathur P and Cordes A W 2003 Inorg. Chim. Acta 353 197; (c) Gupta M, Upadhyay S K, Sridhar M A and Mathur P 2006 Inorg. Chim. Acta 359 4360; (d) Tehlan S, Hundal, M S and Mathur P 2004 Inorg. Chem. 43 6589

- Barnes D J, Chapman R L, Vagg R S and Watton E C 1978 J. Chem. Eng. Data 23 349
- 11. McKee V, Zvagulis M, Dagdigian J V, Patch M G and Reed C A 1984 J. Am. Chem. Soc. **106** 4765
- (a) Tandon S S, Thompson L K, Bridson J N and Dewan J C 1994 *Inorg. Chem.* 33, 54; (b) Addison A W, Burke P J, Henrick K, Rao T N and Sinn E 1983 *Inorg. Chem.* 22 3645; (c) Hendricks H M J, Birker P J M W L, Rijn J V, Verscoor G C and Reedijk J J 1982 *Am. Chem. Soc.* 104 3607; (d) Reitmeijer F J, Birker P J M W L, Gorter S and Reedijk J 1982 *J. Chem. Soc., Dalton Trans.* 1193 (e) Rao T N and Addison A W 1984 *J. Chem. Soc., Dalton Trans.* 1349
- (a) Lloret F, Julve M, Faus J, Journax Y, Levisalles M P and Jeannin Y 1989 Inorg. Chem. 28 3702 (b) Lloret F, Julve M, Faus J, Ruiz R, Castro I, Mollar M and Levisalles M P 1992 Inorg. Chem. 31 784; (c) Frances L, Julve M, Real J, Faus J, Ruiz R, Mollar M, Castro I and Bois C 1992 Inorg. Chem. 31 2956; (d) Lloret F, Sletten J, Ruiz R, Julve M, Faus J and Verdauger M 1992 Inorg. Chem. 31 778 (e) Sangeetha N R, Baradi K, Gupta R, Pal C K, Manivannan V and Pal S 1999 Polyhedron 1999 18 1425
- Vezzosi I M and Antolini I 1984 Inorg. Chim. Acta 85 155
- 15. (a) Prochaska H J, Shwindinger W F, Schwartz M, Burk M J, Bernarducci E, Lalancette R A, Potenza J A and Schugar H J 1981 J. Am. Chem. Soc. 103 3446; (b) Prout C K, Allison G B and Rassottii F J C 1971 J. Chem. Soc. A, 3331; (c) Camerman N, Fawett J K, Kruck T P A, Sarkar B and Camerman A 1978 J. Am. Chem. Soc. 100 2690; (d) Jaffery V, Dagdigian V M and Reed C A 1982 Inorg. Chem. 21 1332
- Thompson L K, Ramaswamy B S and Seymour E A 1977 Can. J. Chem. 55 878
- (a) Monzani E, Battani G, Perotti A, Casella L, Gulloti M, Santagostini L, Nardin G, Randaccio L, Geremia S, Zanello P and Opromolla G 1999 *Inorg. Chem.* 38 5359; (b) Bolus D and Vigee G S 1982 *Inorg. Chim. Acta* 67 19; (c) Zippel F, Ahlers F, Werner R, Haase W, Nolting H F and Krebs B 1996 *Inorg. Chem.* 35 3409
- 18. Cook C D and Norcross B E 1959 J. Org. Chem. 81 1170
- (a) Chin D, La Mar G N and Balch A L 1980 J. Am. Chem. Soc. 102 5945; Balch A L (Private Communication (b) Arasasingham R D, Balch A L and Latos-Grazynski L 1987 J. Am. Chem. Soc. 5846
- 20. Fukuzumi S and Ono Y 1979 Bull. Chem. Soc. Jpn. **52** 2255
- 21. Fukuzumi S and Ono Y 1977 J. Am. Chem. Soc., Perkin Trans. 2 622
- 22. (a) Palaniandavar M, Pandiyan T and Laksminarayanan M 1995 J. Chem. Soc., Dalton Trans. 457 (b) Nonoka Y, Tokh T and Kida S 1974 Bull. Chem. Soc. Jpn. 2 312
- 23. Batra G and Mathur P 1992 Inorg. Chem. 31 1575
- 24. Cook C D and Norcross B E 1958 *J. Am. Chem. Soc.* **81** 1176